



PCRBIO SYSTEMS
simplifying research

2x qPCR BIO Probe Blue Mix Separate-ROX

Product description:

Combined with the latest advancements in polymerase technology and advanced buffer chemistry qPCR BIO Probe Blue Mix offers market leading performance with minimal optimisation.

PCR Biosystems real-time PCR probe mixes have been designed for use on a wide range of probe technologies including TaqMan®, Scorpions® and molecular beacon probes.

qPCR BIO Probe Blue Mix uses antibody-mediated hot start technology that prevents the formation of primer-dimers to improve reaction sensitivity and specificity.

High-throughput screening has resulted in a buffer system that allows efficient amplification from GC-rich and AT-rich templates, under fast and standard cycling conditions.

A blue dye has been added to aid with pipetting. The dye does not interfere with DNA synthesis but will impact the intensity of some fluorescent probes.

Pack size	2x qPCR BIO Probe Blue Mix No-ROX	50µM ROX Additive
100 reactions	1 x 1ml	1 x 200µl
500 reactions	5 x 1ml	1 x 200µl
2000 reactions	20 x 1ml	4 x 200µl
5000 reactions	1 x 50ml	2 x 520µl
5000 reactions	50 x 1ml	2 x 520µl

Shipping and storage

On arrival the kit should be stored at -20°C. Avoid prolonged exposure to light. If stored correctly the kit will retain full activity for 12 months. The kit can be stored at 4°C for 1 month. The kit can go through 30 freeze/thaw cycles with no loss of activity.

Limitations of product use

The product may be used only for in vitro research purposes.

Technical support

For technical support and troubleshooting please email technical@pcrbio.com the following information:

- Amplicon size
- Reaction setup
- Cycling conditions
- Screen grabs of amplification traces and melting profile

www.pcrbio.com

Important considerations

Primer design: For efficient amplification under fast cycling conditions we recommend amplicon lengths between 80bp and 200bp. With all manufacturers' master mixes the shorter the amplicon length the faster the reaction can be cycled. Amplicon lengths should not exceed 400bp. Primers should have a predicted melting temperature of around 60°C, using default Primer 3 settings (<http://frodo.wi.mit.edu/primer3/>). For TaqMan® probes choose probe close to 5' primer, avoid terminal guanosine residues.

Template amount: For genomic DNA, 1µg or less is recommended. For cDNA, 100ng or less is recommended. However, users are encouraged to attempt a dilution series for new template/primer pairs to ensure that the PCR is efficient at that template concentration.

Probe Intensity: qPCRBI Probe Blue Mix will necessarily lower the fluorescent intensity from probes by absorbing light at both the excitation and emission wavelengths (see Table 1). However, the recommended probe concentration of 200nM has proven sufficient for detection on all instruments tested. If signal intensity is a concern, consider switching to a qPCRBI Probe Mix without dye.

Table 1: Fluorescent intensity of selected probes in qPCRBI Probe Blue Mix.

Fluorophore	Ex / Em (nm)	Signal loss
FAM	494 / 518	12%
HEX	535 / 556	55%
Texas Red	595 / 615	88%
Cy5	675 / 694	82%

Instrument compatibility: Different real-time PCR instruments require different levels of ROX passive reference. Generally, modern instruments do not require passive reference but include the option to use it for normalisation. Please check our qPCRBI Selection Table to determine which ROX concentration your instrument requires (<http://www.pcrbio.com/realtime-pcr.html>).

ROX additive protocol: The 50µM ROX Additive supplied is formulated to be added directly to the 1ml tube of 2x qPCRBI master mix supplied. Once the ROX is added, the reagent may be used straight away or stored at -20°C for future use. Please use the following charts to add the correct amount of ROX for your instrument. Vortex thoroughly after ROX addition.

Reagent	Hi-ROX instruments	Lo-ROX instruments
2x qPCRBI Probe Blue Mix No-ROX	1.0ml	1.0ml
50µM ROX Additive	35.0µl	4.0µl

Reaction setup

1. Before starting, briefly vortex 2x qPCRBI Probe Blue Mix.
2. Prepare a master mix based on the following table:

Reagent	20µl reaction	Final concentration	Notes
2x qPCRBI Probe Blue Mix	10µl	1x	
Forward primer (10µM)	0.8µl	400nM	See above for optimal primer design
Reverse primer (10µM)	0.8µl	400nM	
Probe (10µM)	0.4µl	200nM	See above for probe considerations
Template DNA	<100ng cDNA, <1µg genomic	Variable	See above for template considerations
PCR grade dH ₂ O	Up to 20µl final volume		

3. Program the instrument using the following conditions, acquiring data on the appropriate channel:

Cycles	Temperature	Time	Notes
1	95°C	2min	Polymerase activation, 2 minutes for cDNA and 3 minutes for genomic
40	95°C 60°C to 65°C	5 seconds 20-30 seconds	Denaturation Anneal/Extension, do not exceed 30 seconds, do not use temperatures below 60°C
Melt analysis	Refer to instrument instructions		Optional melt profile analysis, available for hybridisation probes only